Reproductive BioMedicine Online (2013) xxx, xxx-xxx



www.sciencedirect.com www.rbmonline.com



ARTICLE

Association between blastocyst morphology and outcome of single-blastocyst transfer

⁵ Etienne Van den Abbeel^a, Basak Balaban^b, Søren Ziebe^c, Kersti Lundin^d,

Maria José Gómez Cuesta ^e, Bjarke Mirner Klein ^f, Lisbeth Helmgaard ^g,
 Joan-Carles Arce ^{g,*}

^a Reproductive Medicine, Gent University Hospital, Gent, Belgium; ^b IVF Center, American Hospital, Istanbul, Turkey;

⁹ ^c The Fertility Clinic, Rigshospitalet, Copenhagen, Denmark; ^d Reproductive Medicine, Sahlgrenska University Hospital,

10 Gothenburg, Sweden; ^e Woman's Health Dexeus, USP Institut Universitari Dexeus, Barcelona, Spain; ^f Global Biometrics,

- Ferring Pharmaceuticals A/S, Copenhagen, Denmark; ^g Reproductive Health, Ferring Pharmaceuticals A/S, Copenhagen,
 Denmark
- ¹³ * Corresponding author. *E-mail address:* jca@ferring.com (J-C Arce).



Etienne Van den Abbeel has a MSc in chemical engineering and a PhD in medical sciences. After spending more than 25 years at the University Hospital of the Dutch-speaking Brussels CRM as a senior clinical embryologist, he is now IVF Laboratory Director at the Centre of Reproductive Medicine of the University Hospital of Ghent, Belgium and professor in clinical embryology at the University of Ghent. He has authored or co-authored over 60 peer-reviewed papers and book chapters. His main research interests are cryobiology and cryopreservation, culture and selection of mammalian gametes and embryos.

Abstract The aim of this study was to assess the ability of three individual blastocyst morphology parameters – expansion and hatching (EH) stage, inner cell mass (ICM) grade and trophectoderm grade – to predict outcome of a cycle with single-blastocyst transfer. The study was a secondary analysis of data prospectively collected in a large multicentre trial. A total of 618 intracytoplasmic sperm injection patients undergoing ovarian stimulation in a gonadotrophin-releasing hormone antagonist cycle with compulsory single-blastocyst transfer on day 5 were included. In the simple logistic regression analysis, all three blastocyst morphology parameters were statistically significantly (P < 0.005 for each) associated with positive human chorionic gonadotrophin, clinical and ongoing pregnancy rates and live birth rates, while only the ICM grade was significantly (P = 0.033) associated with early pregnancy

Q1 loss rate. Blastocyst EH stage was the only significant predictor of live birth (P < 0.001) in the multiple logistic regression. In conclusion, although all three blastocyst morphology parameters were related to treatment outcome of fresh single-blastocyst cycles, selection of high-quality blastocysts for transfer should consider first the EH stage. Transfer of a blastocyst with ICM grade A may reduce the risk of early pregnancy loss.

© 2013, Reproductive Healthcare Ltd. Published by Elsevier Ltd. All rights reserved.

11 KEYWORDS: blastocyst morphology, early pregnancy loss, live birth, prediction, single-embryo transfer

1472-6483/\$ - see front matter © 2013, Reproductive Healthcare Ltd. Published by Elsevier Ltd. All rights reserved. http://dx.doi.org/10.1016/j.rbmo.2013.07.006

121

12 Introduction

13 Weighing the benefits and risks of assisted reproduction 14 technology, the birth of a single healthy child is the ultimate objective in patients undergoing ovarian stimulation 15 for IVF/intracytoplasmic sperm injection (ICSI) (Land and 16 17 Evers, 2003). Single-embryo transfer would appear to be the most effective approach to ensure this objective. The 18 advantages of reducing multiple pregnancies in terms of 19 obstetric risk and child health outcome are obvious, but 20 21 the impact on the overall efficacy of single- versus 22 multiple-embryo transfer has been debated (Bergh, 2005; 23 Gelbava et al., 2010; Martikainen et al., 2001; McLernon 24 et al., 2010; Pandian et al., 2009; Thurin et al., 2004). Systematic reviews have concluded that while single-embryo 25 transfer is associated with lower live birth rate than dou-26 27 ble-embryo transfer in fresh cycles, the cumulative live 28 birth rates from fresh and frozen single-embryo transfer 29 cycles are similar to those in patients undergoing fresh double-embryo transfer (Gelbaya et al., 2010; McLernon et al., 30 31 2010; Pandian et al., 2009).

Embryo quality is considered a major predictor of 32 implantation and pregnancy (Ahlström et al., 2011; della 33 34 Ragione et al., 2007; Richter et al., 2001; Terriou et al., 2001; Thurin et al., 2005; Van Royen et al., 1999; Ziebe 35 36 et al., 1997). Selecting the embryo(s) with the best implan-37 tation potential is essential for securing each couple the 38 highest chance of achieving pregnancy after assisted repro-39 duction. The ability to make the optimal choice has become 40 even more important with the growing implementation of single-embryo transfers. In the early 1990s, knowledge of 41 42 metabolic requirements of the developing embryo increased 43 and sequential embryo culture media were introduced (Gardner and Lane, 1999). This rapidly increased the pro-44 45 portion of embryos developing to the blastocyst stage. 46 Extended embryo culture to the blastocyst stage has 47 allowed for assessment of embryo morphology beyond geno-48 mic activation and thus contributed to improved embryo 49 selection. There is increasing evidence that an embryo's 50 ability to reach the blastocyst stage in vitro improves pre-51 diction of clinical pregnancy (Blake et al., 2007; Rehman et al., 2007) and that transfer of blastocysts results in 52 53 higher live birth rates than those achieved with the same number of cleavage-stage embryos (Papanikolaou et al., 54 55 2006, 2008).

A number of classification and grading systems are avail-56 57 able for assessing embryo quality at the cleavage stage (Alpha Scientists in Reproductive Medicine and ESHRE Spe-58 59 cial Interest Group of Embryology, 2011; Cummins et al., 1986; Fisch et al., 2001; Giorgetti et al., 1995; Holte 60 et al., 2007; Prados et al., 2012; Puissant et al., 1987; Ter-61 riou et al., 2001; Van Royen et al., 1999), but only a few 62 63 grading systems have been proposed for evaluating quality 64 at the blastocyst stage. In 1999, Gardner and Schoolcraft 65 introduced a blastocyst grading system in which selection 66 of high-quality blastocysts is based on morphology parameters of the expansion and hatching (EH) stage, the inner cell 67 mass (ICM) grade and the trophectoderm (TE) grade (Gard-68 ner and Schoolcraft, 1999). Composite scores of the differ-69 70 ent morphology parameters (Balaban et al., 2006; della 71 Ragione et al., 2007; Gardner et al., 2000; Goto et al., 2011)

have been linked to pregnancy and/or pregnancy loss. Other 72 quantitative measures of blastocyst morphology and their 73 relation to implantation and pregnancy have been evaluated 74 (Richter et al., 2001; Shapiro et al., 2008), but the blasto-75 cyst grading system by Gardner and Schoolcraft (1999) 76 remains largely unchallenged. There is, however, a need 77 for increased knowledge of the relative impact of each mor-78 phology parameter at the blastocyst stage, as well as their 79 correlations, in predicting the probability of successful 80 implantation and pregnancy. Such data would be most reli-81 able when based on single-blastocyst transfers; however, 82 the majority of the available data on the association 83 between blastocyst quality and outcome parameters have 84 been derived from studies using multiple-blastocyst trans-85 fers and, so far, only a few studies reporting data from fresh 86 single-blastocyst transfers exist (Ahlström et al., 2011; della 87 Ragione et al., 2007; Hill et al., 2013; Kresowik et al., 88 2012). 89

The main aim of this study was to investigate the rela-90 tionship between individual morphology parameters and 91 pregnancy, pregnancy loss and live birth, using data 92 obtained from a large multicentre trial with compulsory sin-93 gle-blastocyst transfer (Devroey et al., 2012). A secondary 94 aim was to develop a multiple logistic regression model 95 for prediction of the probability of live birth after sin-96 gle-blastocyst transfer based on blastocyst morphology 97 scoring. 98

Materials and methods

This study is a secondary analysis of data prospectively col-100 lected from all patients (n = 618) who had compulsory sin-101 gle-blastocyst transfer on day 5 after oocyte retrieval 102 while participating in a multicentre randomized controlled 103 trial comparing ongoing pregnancy rates after ovarian stim-104 ulation with highly purified human menopausal gonadotro-105 phin (HP-HMG; Menopur; Ferring Pharmaceuticals) or 106 recombinant FSH (follitropin β ; Puregon; MSD) in ICSI 107 patients following a gonadotrophin-releasing hormone 108 (GnRH) antagonist cycle. The trial was carried out in accor-109 dance with the declaration of Helsinki, International Con-110 ference on Harmonization Guidelines for Good Clinical 111 Practice and local regulatory requirements. The trial was 112 registered at ClinicalTrials.gov (number NCT00884221) and 113 the protocol (FE 999906 CS08) was approved by the local 114 regulatory authorities and the independent ethics commit-115 tees covering all participating centres. Written informed 116 consent was provided by all patients before any trial-related 117 examinations were initiated. The trial design, population, 118 methods, conduct and results have been reported previously 119 (Devroey et al., 2012). 120

Trial population

The main inclusion criteria were women with primary diagnosis of infertility being unexplained infertility or partners122nosis of infertility being unexplained infertility or partners123with mild male factor, age 21–34 years, body mass index12418–25 kg/m², FSH 1–12 IU/l, antral follicle count \geq 10 and125regular menstrual cycles of 24–35 days. Women with poly-126cystic ovaries, endometriosis stage I–IV or poor response127in a previous stimulation cycle were excluded.128

Treatment regimen 129

130 The starting gonadotrophin dose was fixed at 150 IU daily for the first 5 days and adjusted according to ovarian response 131 from day 6 when the GnRH antagonist (ganirelix acetate; 132 Orgalutran; MSD) was initiated at a daily dose of 0.25 mg 133 and continued throughout the gonadotrophin-treatment 134 135 period. A single injection of 250 µg HCG (Ovitrelle; Merck 136 Serono) was administered as soon as three follicles of >17 mm were observed. Oocyte retrieval took place 137 138 36 ± 2 h after the HCG administration.

Blastocyst assessments 139

All oocytes retrieved were fertilized by ICSI. Fertilization 140 was assessed $19 \pm 1 h$ post insemination and embryos with 141 142 two pronuclei were cultured individually (in separate droplets) and assessed daily by the local embryologists. Only 143 144 commercially available culture media were used from 145Q2 retrieval to transfer. On day 5 (120 \pm 2 h) post insemination, 146 the blastocyst quality was assessed based on morphological criteria and a single blastocyst of the best quality, according 147 to the local embryologist, was transferred. Remaining blas-148 149 tocysts were cryopreserved individually by vitrification.

The morphological evaluation of blastocysts was per-150 formed according to the Gardner and Schoolcraft grading 151 system (Gardner and Schoolcraft, 1999) and included three 152 different parameters: EH stage, ICM grade and TE grade. 153 The EH stage was assessed as one of the following: (1) an 154 155 early blastocyst, blastocoele being less than half volume 156 of that of the embryo; (2) a blastocyst with a blastocoele whose volume is half of, or greater than half of that of 157 the embryo; (3) a full blastocyst with a blastocoele com-158 159 pletely filling the embryo; (4) an expanded blastocyst with a blastocoele volume larger than that of the full blastocyst, 160 161 with a thinning zona; (5) a hatching blastocyst with the TE starting to herniate through the zona; and (6) a hatched 162 163 blastocyst, in which the blastocyst has completely escaped from the zona. For blastocysts with EH stage >3, ICM grade 164 and TE grade were evaluated. The ICM was assessed as one 165 of the following: (A) tightly packed, many cells; (B) loosely 166 grouped, several cells; and (C) very few cells. The TE was 167 assessed as one of the following: (A) many cells forming a 168 cohesive epithelium; (B) few cells forming a loose epithe-169 170 lium; and (C) very few, large cells. To harmonize intra-171 and interclinic scoring of the morphology parameters, a 172 common prestudy training session was held with the respon-173 sible embryologist(s) from each of the participating centres. 174 All embryologists had also to pass an online scoring test provided by Fertaid (www.fertaid.com) to be able to partici-175 pate in the trial. In addition, an atlas with representative 176 177 pictures of all morphology parameters was prepared as a visual aid and distributed to all embryologists before the 178 start of the trial to further ensure standardized assess-179 180 ments. Examples of blastocyst grading included in the atlas are shown in Figure 1. 181

Clinical outcome 182

183 A serum β HCG test was performed 13–15 days after blastocyst transfer. Clinical and ongoing pregnancy was 184 185 confirmed by transvaginal ultrasound 5-6 and 10-11 weeks, respectively, after transfer. Early pregnancy loss was 186 defined as a pregnancy loss occurring between the positive β HCG test and ongoing pregnancy, and late pregnancy loss as a pregnancy loss occurring after confirmed ongoing pregnancy. All patients with an established ongoing pregnancy 190 were followed until delivery. 191

Statistical analysis

Continuous data were presented as median values with 193 interguartile range (IQR) with differences between groups 194 tested using Wilcoxon's test. Categorical data were pre-195 sented as frequencies and percentages accompanied with 196 P-values based on the likelihood ratio chi-squared test. All 197 reported P-values were two-sided. A P-value < 0.05 was 198 considered significant. No adjustment for multiplicity was 199 applied. The interdependency between the blastocyst grade 200 parameters was evaluated using pairwise chi-squared tests. 201 Assessment of the relationship between clinical outcome 202 203 (positive β HCG, early pregnancy loss, clinical and ongoing pregnancy, and live birth) and the individual grading param-204 eters and potential confounding parameters were based on 205 logistic regression. The following strategy was applied: each 206 parameter was analysed using simple logistic regression to 207 identify potential candidates for the multiple logistic 208 regression modelling. Parameters identified as significant 209 in the simple logistic regression were entered in a multiple 210 logistic regression model of live birth rate. Stepwise back-211 ward elimination was then performed: i.e. the least signifi-212 cant parameter was removed (eliminated) and the model 213 was then refitted. This process was repeated until all 214 remaining variables were significant. Based on the final 215 model, the predicted probabilities of live birth accompa-216 nied with 95% Wald confidence limits were reported. 217

Results

A total of 618 women (HP-HMG n = 304; recombinant FSH 219 n = 314) underwent a compulsory single-blastocyst transfer 220 on day 5 after oocyte retrieval. For this investigation, data 221 from all patients were integrated as no differences were 222 observed between HP-HMG and recombinant FSH groups 223 on baseline characteristics, end-of-stimulation ovarian 224 response, number of blastocysts available on day 5, distribu-225 tion of EH stage. ICM and TE for transferred blastocysts and 226 live birth rates. Table 1 shows the composite grading of the 227 transferred blastocysts. The three morphology parameters 228 were pairwise positively associated (P < 0.001), with EH 229 stage 5 more frequently associated with ICM and TE grades 230 AA, stage 4 with grades AA or BB and stage 3 with grades BB 231 or CC. No blastocysts with EH stage 6 were observed for any 232 of the patients. 233

Treatment outcome according to blastocyst quality 234

There were no differences between the patients who 235 achieved a live birth (n = 200, 32%) and those who did not 236 (n = 418, 68%) concerning patient characteristics, except 237 for a significantly lower serum progesterone concentration 238 (P = 0.047) and a higher serum LH concentration (P = 0.028)239 at the end of stimulation in patients with live birth 240 (Table 2). The patients who achieved a live birth had signif-241

Please cite this article in press as: Van den Abbeel, E et al. Association between blastocyst morphology and outcome of single-blastocyst transfer. Reproductive BioMedicine Online (2013), http://dx.doi.org/10.1016/j.rbmo.2013.07.006

3

187 ς

189

192

218

E Van den Abbeel et al.



Figure 1 Examples of blastocyst grading: (A) 3AA blastocyst; (B) 3AB blastocyst; (C) 3BA blastocyst; (D) 4AA blastocyst; (E) 4AB blastocyst; (F) 4BA blastocyst; (G) 4CC blastocyst; (H) 5AA blastocyst; (I) 5CA blastocyst. For details of the EH stages and ICM and TE grades, see Materials and methods. Bars = $50 \mu m$.

Table 1	Distribution of	composite	morphology	parameters of	transferred	blastocysts	on day !	5
post inser	nination.							

EH stage	n	ICM and TE grades								
		AA	АВ	AC	BA	BB	ВС	СА	СВ	СС
6	0	_		-	_	_	_	_	_	_
5	152	80 (53)	28 (18)	0 (0)	15 (10)	22 (14)	5 (3)	0 (0)	0 (0)	2 (1)
4	255	101 (40)	38 (15)	1 (<1)	20 (8)	63 (25)	7 (3)	3 (1)	12 (5)	10 (4)
3	106	9 (8)	10 (9)	3 (3)	6 (6)	37 (35)	9 (8)	1 (1)	7 (7)	24 (23)
2 ^a	52	_	_	_	_	_	_	_	_	_
1 ^a	53	-	-	-	_	_	_	-	-	-

Values are *n* (%).

EH = expansion and hatching; ICM = inner cell mass; TE = trophectoderm. a ICM and TE grades were not evaluated.

icantly more blastocysts available on day 5: median (IQR) 3 242 243 (2-5) versus 2 (1-4); P < 0.001. The relative distribution of individual scores for each morphology parameter for the 244 245 transferred blastocysts were significantly different between 246 the patients with a live birth and those with no live birth (P < 0.001 for each parameter). For patients with a live 247 birth, a larger proportion of the transferred blastocysts 248 249 were of EH stages 4 or higher (87% versus 56%), ICM grade A (62% versus 47%) and TE grade A (55% versus 44%) com-250 251 pared with the patients with no live birth (Table 2).

In the simple logistic regression analysis, increasing blas-252 tocyst EH stage was positively associated with positive 253 β HCG, clinical pregnancy, ongoing pregnancy and live birth (P < 0.001 for each). Female age, body mass index, primary 255 cause of infertility, type of gonadotrophin preparation and 256 serum concentrations of FSH, LH, oestradiol and progester-257 one at end of stimulation were not significantly associated 258 with any parameter of treatment outcome. Table 3 displays 259 the observed positive β HCG, clinical pregnancy, ongoing 260 pregnancy and live birth rates as well as the early pregnancy 261

Blastocyst morphology and outcome of single-blastocyst transfer

Characteristic	No live birth (n = 418)	<i>Live birth (</i> n = 200)	P-value ^b
Baseline			
Female age (years)	31 (29-33)	31 (29-33)	NS
Body mass index (kg/m ²)	21.8 (20.3–23.6)	22.0 (20.4–23.5)	NS
Primary cause of infertility			NS
Unexplained infertility	38	40	
Mild male factor	62	60	
No. of previous ovarian stimulation cycles	0 (0–1)	0 (0–0)	NS
Day 1 (before start of stimulation)			
Antral follicle count	15 (12–18)	15 (12–19)	NS
Anti-Müllerian hormone (pmol/l)	23 (13-38)	26 (14-38)	NS
LH (IU/l)	5.8 (4.6-7.4)	5.9 (4.7–7.6)	NS
FSH (IU/l)	6.9 (6.1–8.2)	7.0 (6.2-8.1)	NS
Endometrial thickness (mm)	3 (2-5)	3 (3–5)	NS
End of stimulation			
Oestradiol (nmol/l)	6.6 (4.6–9.5)	6.2 (4.6-9.4)	NS
Progesterone (nmol/l)	2.6 (1.9–3.5)	2.4 (1.8–3.4)	0.047
LH (IU/l)	1.8 (1.0–2.8)	2.0 (1.2–3.2)	0.028
Endometrial thickness (mm)	11 (10–12)	11 (9–12)	NS
Total gonadotrophin dose (IU)	1350 (1200–1500)	1350 (1200–1500)	NS
No. of oocytes retrieved	9 (6–13)	9 (6–14)	NS
Blastocysts available on day 5	2 (1-4)	3 (2-5)	<0.001
EH stage			<0.001
5	82 (20)	70 (35)	
4	152 (36)	103 (52)	
3	88 (21)	18 (9)	
2	46 (11)	6 (3)	
1	50 (12)	3 (2)	
ICM grade ^a			<0.001
Α	151 (47)	119 (62)	
В	124 (39)	60 (31)	
с	47 (15)	12 (6)	
TE grade ^a			<0.001
Ă	130 (40)	105 (55)	
В	143 (44)	74 (39)	
c L	49 (15)	12 (6)	
-	. (,		

 Table 2
 Patient and morphology characteristics of transferred blastocysts by live birth status.

Values are median (interquartile range), % or n (%).

EH = expansion and hatching; ICM = inner cell mass; TE = trophectoderm.

^aFor blastocysts of EH stage 3-5.

^bWilcoxon's test (continuous data) or the likelihood ratio chi-squared test (categorical data).

loss rates according to the individual grades of the three 262 263 blastocyst morphology parameters. The live birth rate 264 increased from 6% for transfer of EH stage 1 blastocysts to 46% for EH stage 5 blastocysts. For blastocysts with EH 265 stages 3-5, the ICM grade had a significant impact on the 266 likelihood of achieving a positive β HCG (*P* = 0.004), clinical 267 pregnancy (P = 0.002), ongoing pregnancy (P < 0.001) and 268 269 live birth (P < 0.001). Likewise, the TE grade showed signif-270 icant predictive value for positive β HCG (*P* = 0.002), clinical pregnancy (P = 0.003), ongoing pregnancy (P = 0.002) and 271 live birth (P < 0.001). The live birth rate increased from 272 20% for ICM grade C to 44% with grade A, and from 20% for 273 TE grade C to 45% with grade A. When restricting the eval-274 uation to blastocysts with EH stages 4 and 5, there were 275 no significant associations between the outcome parame-276 ters and ICM grade or TE grade. Of the three quality param-277 eters, only the ICM grade was found to be significantly 278 (P = 0.033) associated with early pregnancy loss. This associ-279

ARTICLE IN PRESS

E Van den Abbeel et al.

Table 3	Clinical outcome	oy morphology	characteristics of	transferred blastoc	ysts on day 5	post insemination.
---------	------------------	---------------	--------------------	---------------------	---------------	--------------------

Blastocyst grade component	Blastocysts transferred	Positiv	re βHCG	Clinica pregna	Clinical pregnancy		Ongoing pregnancy		Live birth		Early pregnancy loss	
	n <i>(%)</i>	n <i>(%)</i>	P- value	n <i>(%)</i>	P- value	n <i>(%)</i>	P- value	n <i>(%)</i>	P- value	n <i>(%)</i>	P- value	
EH stage												
All data (<i>n</i> = 618) 5	152 (25)	93 (61)	<0.001	76 (50)	<0.001	72 (47)	<0.001	70 (46)	<0.001	21 (23)	NS	
4	255 (41)	139 (55)		(45)		106 (42)		103 (40)		33 (24)		
3	106 (17)	27 (25)		21 (20)		19 (18)		18 (17)		8 (30)		
2	52 (8)	13 (25)		6 (12)		6 (12)		6 (12)		7 (54)		
1	53 (9)	5 (9)		3 (6)		3 (6)		3 (6)		2 (40)		
ICM grade All data (n = 513)												
Α	270 (53)	150 (56)	0.004	130 (48)	0.002	123 (46)	<0.001	119 (44)	<0.001	27 (18)	0.033	
В	184 (36)	90 (49)		66 (36)		61 (33)		60 (33)		29 (32)		
C	59 (12)	19 (32)		16 (27)		13 (22)		12 (20)		6 (32)		
Blastocysts with EH stage 4 or 5 (n = 407)												
A	248 (61)	144 (58)	NS	124 (50)	NS	117 (47)	NS	115 (46)	NS	27 (19)	NS	
В	132 (32)	73 (55)		54 (41)		49 (37)		48 (36)		24 (33)		
С	27 (7)	15 (56)		13 (48)		12 (44)		11 (41)		3 (20)		
<i>TE grade</i> All data (<i>n</i> = 513)												
А	235 (46)	135 (57)	0.002	112 (48)	0.003	107 (46)	0.002	105 (45)	<0.001	28 (21)	NS	
В	217 (42)	104 (48)		85 (39)		76 (35)		74 (34)		28 (27)		
С	61 (12)	20 (33)		15 (25)		14 (23)		12 (20)		6 (30)		
Blastocysts with EH stage 4 or 5 (n = 407)	X											
A	219 (54)	131	NS	110	NS	104	NS	103	NS	27	NS	
В	163 (40)	(00) 88 (54)		(50) 73 (45)		(47) 65 (40)		(47) 64 (39)		(21) 23 (26)		
С	25 (6)	13 (52)		9 (36)		9 (36)		7 (28)		4 (31)		

Simple logistic regression.

EH = expansion and hatching; ICM = inner cell mass; TE = trophectoderm.

No. of Pages 9, Model 6+

7

306

ation did not remain significant when restricting the analysis
to blastocysts with EH stages 4 and 5. In total, there were
only six pregnancy losses after confirmed ongoing pregnancy
in the study and no evaluation of the association between
blastocyst morphology and late pregnancy loss was
performed.

286 **Prediction of live birth rate**

Only the blastocyst quality parameters were included in the 287 288 multiple logistic regression analysis of live birth rate, as no 289 confounding parameters were significantly associated with 290 live birth rate in the simple logistic regression analyses. 291 Stages 4 and 5 of blastocyst EH were combined in the pre-292 diction model because there was no significant difference 293 with respect to live birth rate between these two stages. 294 After stepwise backward elimination, the minimal model 295 simply included blastocyst EH stage, as the predictive power (P = 0.002) of this parameter overruled those of the other 296 two parameters. However, because of the strong signifi-297 298 cance of ICM and TE grade observed in the simple logistic 299 regression, these were reintroduced in the final model.

Table 4 provides the estimated probabilities of achieving
a live birth depending on the composite quality score of the
blastocyst. As examples for the model, a patient with transfer of a blastocyst with a score of 4–5AA, 4–5BB or 3CC is
predicted to have a probability of achieving a live birth of
47%, 38% or 12%, respectively.

Table 4Predicted live birth rate by compositemorphology classification of transferred blastocysts.

EH stage	ICM grade	TE grade	Live birth rate
4 or 5	A A B B C C C	A B C A B C A B C	47 (41–54) 43 (33–53) 37 (20–58) 42 (32–53) 38 (30–47) 32 (17–51) 36 (19–56) 32 (17–51) 27 (14–45)
3	A A B B C C C C	A B C A B C A B C	24 (15–38) 21 (12–34) 17 (8–35) 21 (12–34) 18 (11–28) 15 (7–29) 17 (7–34) 14 (6–29) 12 (5–23)
2	-	_	12 (5–23)
1	-	-	6 (2-16)

Values are % (95% CI). Predicted live birth rate based on the multiple logistic regression model.

EH = expansion and hatching; ICM = inner cell mass; TE = trophectoderm.

Discussion

The present study in a single-blastocyst setting showed that 307 high scores of blastocyst EH stage, ICM grade and TE grade 308 were all significantly associated with increased pregnancy 309 and live birth rates after fresh transfers. The relevance of 310 blastocyst EH stage has been previously documented, as 311 higher implantation rates were obtained with transfer of 312 expanded blastocysts compared with non-expanded blasto-313 cysts (della Ragione et al., 2007; Kresowik et al., 2012; 314 Racowsky et al., 2003; Wilson et al., 2004) and with hatch-315 ing blastocysts compared with non-hatching blastocysts 316 (Balaban et al., 2000; Yoon et al., 2001). Previous studies 317 have also reported that an ICM tightly packed with many 318 cells contributes to vital implantation or live birth rate 319 (Kovacic et al., 2004; Richter et al., 2001). In the present 320 study, ICM was positively associated with BHCG, clinical 321 and ongoing pregnancy and live birth rates; interestingly, 322 the percentage of early pregnancy loss after transfer of a 323 blastocyst with ICM grade A was about half of that after 324 transfer of a blastocyst with ICM grade B or C suggesting that 325 a large ICM increases the probability of maintaining the 326 pregnancy beyond the initial positive BHCG finding, reflect-327 ing a higher viability of the initial implantation. 328

When including all three morphology parameters in a 329 logistic regression model of live birth rate, only the EH stage 330 remained as a significant independent predictor. However, 331 because of the strong significance of ICM and TE grade 332 observed in the simple logistic regression, a composite clas-333 sification consisting of all three morphology parameters 334 were used in the model for prediction of live birth. Thus, 335 transfer of a blastocyst with a high stage (4 or 5) of EH 336 and high grades of ICM and TE (i.e. AA) was estimated to 337 result in approximately twice as high chance of obtaining 338 a live birth compared with transfer of low grades of ICM 339 and TE (i.e. CC). It should be noted that the actual pre-340 dicted probabilities of obtaining a live birth could not be 341 discriminated in the present data set in which the 95% con-342 fidence intervals overlapped. 343

The finding in the present study that the EH stage is the 344 most important parameter when selecting a blastocyst for 345 transfer is in contrast with some recent retrospective cohort 346 studies suggesting the TE grade to have the strongest pre-347 dictive power for treatment outcome in fresh transfers 348 (Ahlström et al., 2011; Hill et al., 2013) and frozen-thawed 349 transfers (Honnma et al., 2012). Although these studies also 350 used the grading system of Gardner and Schoolcraft (1999), 351 differences in the relative distribution of blastocysts with 352 different morphology grades between the studies may con-353 tribute to explain the apparently discrepant relative impor-354 tance of blastocyst morphology parameters: the vast 355 majority of blastocysts included were of good or excellent 356 quality, with very few (5% or less) blastocysts with C grades 357 for ICM or TE; however, the data set in present study was 358 derived from a clinical trial in which single-embryo transfer 359 was mandatory irrespective of the blastocyst guality and 360 therefore included significant numbers of blastocysts with 361 EH stages 1–5 as well as ICM and TE grades of A, B and C. 362 Furthermore, all data in the present study were derived 363 from only one fresh cycle per patient. The results were also 364 less influenced by confounding parameters, as the study 365

cohort consisted of good-prognosis patients with a narrow 366 367 age range and who were prospectively managed in a harmo-368 nized and standardized manner, i.e. all patients underwent a similar ovarian stimulation protocol and all oocytes were 369 370 fertilized by ICSI and cultured to a similar time point on 371 day 5 (120 \pm 2 h). On the other hand, an inherent problem 372 with the blastocyst grading system introduced by Gardner 373 and Schoolcraft (1999) is the loose definitions used of the 374 ICM and TE morphology. Despite written definitions and a 375 visual aid atlas as well as common training sessions, it cannot be ruled out that the blastocyst scoring was still to some 376 377 extent a subjective assessment.

378 In the present study, ICM and TE A and B grades were 379 more frequently associated with higher blastocyst EH 380 stages. Also, the qualities of ICM and TE were highly associated, i.e. AA, BB or CC grades occurred more frequently 381 382 than other possible combinations. These observations may 383 indicate interdependency between developmental stage, ICM and TE of the blastocyst, increasing the complexity 384 when interpreting the actual impact of the individual fac-385 386 tors. A better understanding of the correlation and potential 387 interlink between these three morphology parameters may 388 lead to a more consensus-oriented position in the selection 389 of the best blastocyst for transfer. As discussed by Ahlström 390 et al. (2011), the extent of blastocyst expansion is related 391 to the number and cohesiveness of the TE cells, which would prevent leakage of the blastocoele liquid and sodium 392 393 ions. Thus, a good TE grade may reflect that the blastocyst 394 is efficiently pumping ions into the cavity and inducing 395 osmotic accumulation of water in the cells resulting in 396 higher blastocyst expansion (Ahlström et al., 2011). In other 397 words, a fully expanded blastocyst by definition requires a 398 functional TE, and this functionality may be more depen-399 dent on the molecular guality of TE cells than on their guan-400 tity and cohesiveness.

401 Finally, other quantitative aspects of blastocyst expan-402 sion. ICM and TE not currently evaluated may play a role 403 in blastocyst implantation. In this respect, other measures 404 of ICM such as size, shape and fragmentation, or measures 405 of cell number, blastocyst diameter and blastulation timing, have been reported to be associated with implantation 406 407 potential and viability (Richter et al., 2001; Shapiro et al., 408 2008). Further research is needed to evaluate whether addi-409 tional blastocyst parameters, or embryo development 410 parameters at earlier time points, could contribute to provide a more precise prediction model of live birth based 411 on single-blastocyst transfer in fresh cycles. 412

In conclusion, the EH stage should be considered first
among the three morphology parameters when selecting a
blastocyst for transfer, as this parameter has the highest
predictive value of live birth. At any blastocyst EH stage,
additional consideration should be given to both ICM and
TE grade. Transfer of a blastocyst with ICM grade A may
reduce the risk of an early pregnancy loss.

420 Acknowledgements

The authors thank Göran Pettersson, PhD, Reproductive
Health, Ferring Pharmaceuticals for assistance in writing
the manuscript. The authors also thank all staff at the
participating centres: (i) Belgium: Universitair Ziekenhuis

Brussel, Brussels; Hôpital Erasme, Brussels; Universitair Zie-425 kenhuis Gent, Gent; Universitair Ziekenhuis Antwerpen, 426 Edegem; (ii) Czech Republic: ISCARE IVF, Prague; IVF Insti-427 tute, Pilsen; Pronatal, Prague; (iii) Denmark: H:S Rigshospi-428 Copenhagen; Sygehus Vestsjælland, talet. Holbæk: 429 Amtssygehuset, Herlev; H:S Hvidovre Hospital, Hvidovre; 430 (iv) Poland – KRIOBANK, Bialystok; nOvum, Warsaw; (v) 431 Spain: GINEFIV, Madrid; IU Dexeus, Barcelona; IVI Madrid, 432 Madrid; Ginemed Sevilla, Sevilla; IVI Sevilla, Sevilla; IVI 433 Valencia, Valencia; (vi) Sweden: IVF-kliniken CURA, Malmö; 434 Fertilitetscentrum AB, Gothenburg; RMC, Malmö; (vii) Tur-435 key: Hacettepe University, Ankara, American Hospital, 436 Istanbul; Memorial Hospital, Istanbul. 437

References

- Ahlström, A., Westin, C., Reismer, E., Wikland, M., Hardarson, T., 2011. Trophectoderm morphology: an important parameter for predicting live birth after single blastocyst transfer. Hum. Reprod. 26, 3289–3296.
- Alpha Scientists in Reproductive Medicine and ESHRE Special Interest Group of Embryology, 2011. The Istanbul consensus workshop on embryo assessment: proceedings of an expert meeting. Hum. Reprod. 26, 1270–1283.
- Balaban, B., Yakin, K., Urman, B., 2006. Randomized comparison of two different blastocyst grading systems. Fertil. Steril. 85, 559–563.
- Balaban, B., Urman, B., Sertac, A., Alatas, C., Aksoy, S., Mercan, R., 2000. Blastocyst quality affects the success of blastocyst-stage embryo transfer. Fertil. Steril. 74, 282–287.
- Bergh, C., 2005. Single embryo transfer: a mini-review. Hum. Reprod. 20, 323–327.
- Blake, D.A., Farquhar, C.M., Johnson, N., Proctor, M., 2007. Cleavage stage versus blastocyst stage embryo transfer in assisted conception. Cochrane Database Syst. Rev., CD002118.
- Cummins, J.M., Breen, T.M., Harrison, K.L., Shaw, J.M., Wilson, L.M., Hennessey, J.F., 1986. A formula for scoring human embryo growth rates in in vitro fertilization: its value in predicting pregnancy and in comparison with visual estimates of embryo quality. J. In Vitro Fert. Embryo Transf. 3, 284–295.
- della Ragione, T., Verheyen, G., Papanikolaou, E.G., Van Landuyt, L., Devroey, P., Van Steirteghem, A., 2007. Developmental stage on day-5 and fragmentation rate on day-3 can influence the implantation potential of top-quality blastocysts in IVF cycles with single embryo transfer. Reprod. Biol. Endocrinol. 5, 2.
- Devroey, P., Pellicer, A., Nyboe Andersen, A., Arce, J.-C.on behalf of the Menopur in GnRH Antagonist Cycles with Single Embryo Transfer (MEGASET) Trial Group, 2012. A randomized assessor-blind trial comparing highly purified hMG and recombinant FSH in a GnRH antagonist cycle with compulsory single-blastocyst transfer. Fertil. Steril. 97, 561–571.
- Fisch, J.D., Rodriguez, H., Ross, R., Overby, G., Sher, G., 2001. The graduated embryo score (GES) predicts blastocyst formation and pregnancy rate from cleavage-stage embryos. Hum. Reprod. 16, 1970–1975.
- Gardner, D., Lane, M., 1999. Embryo culture systems. In: Trounson, A.O., Gardner, D.K. (Eds.), Handbook of In Vitro Fertilization, second ed. CRC Press, Boca Raton, FL, USA, pp. 205–264.
- Gardner, D.K., Schoolcraft, W.B., 1999. In-vitro culture of human blastocysts. In: Jansen, R., Mortimer, D. (Eds.), Towards Reproductive Certainty: Fertility and Genetics Beyond 1999. The Parthenon Publishing Group, New York, pp. 378–388.
- Gardner, D.K., Lane, M., Stevens, J., Schlenker, T., Schoolcraft, W.B., 2000. Blastocyst score affects implantation and pregnancy

Please cite this article in press as: Van den Abbeel, E et al. Association between blastocyst morphology and outcome of single-blastocyst transfer. Reproductive BioMedicine Online (2013), http://dx.doi.org/10.1016/j.rbmo.2013.07.006

438 439

440

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

456

457

458

459

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

479

480

481

482

483

484

485

486

487

489

9

548 549

550

551

552

553

554

555 556

557

558

559

560

561

562

563

564

565

566

567

568

569 570

571

572

573

574 575

576

577

578

579

580

581

582

583

584

585

586

587

588

589

590

591

592

593

outcome: towards a single blastocyst transfer. Fertil. Steril. 73, 1155–1158.

Blastocyst morphology and outcome of single-blastocyst transfer

- Gelbaya, T.A., Tsoumpou, I., Nardo, L.G., 2010. The likelihood of
 live birth and multiple birth after single versus double embryo
 transfer at the cleavage stage: a systematic review and
 meta-analysis. Fertil. Steril. 94, 936–945.
- Giorgetti, C., Terriou, P., Auquier, P., Hans, E., Spach, J.L.,
 Salzman, J., Roulier, R., 1995. Embryo score to predict implantation after in vitro fertilization: based on 957 single embryo
 transfers. Hum. Reprod. 10, 2427–2431.
- Goto, S., Kadowaki, T., Tanaka, S., Hashimoto, H., Kokeguchi, S.,
 Shiotani, M., 2011. Prediction of pregnancy rate by blastocyst
 morphological score and age, based on 1,488 single frozen-thawed blastocyst transfer cycles. Fertil. Steril. 95,
 948–952.
- Hill, M.J., Richter, K.S., Heitmann, R.J., Graham, J.R., Tucker,
 M.J., Decherney, A.H., Browne, P.E., Levens, E.D., 2013.
 Trophectoderm grade predicts outcomes of single-blastocyst
 transfers. Fertil. Steril. 99, 1283–1289.
- Holte, J., Berglund, L., Milton, K., Garello, C., Gennarelli, G.,
 Revelli, A., Bergh, T., 2007. Construction of an evidence-based
 integrated morphology cleavage embryo score for implantation
 potential of embryos scored and transferred on day 2 after
 oocyte retrieval. Hum. Reprod. 22, 548–557.
- Honnma, H., Baba, T., Sasaki, M., Hashiba, Y., Ohno, H., Fukunaga,
 T., Endo, T., Saito, T., Asada, Y., 2012. Trophectoderm
 morphology significantly affects the rates of ongoing pregnancy
 and miscarriage in frozen-thawed single-blastocyst transfer
 cycle in vitro fertilization. Fertil. Steril. 98, 361–367.
- 517 Kovacic, B., Vlaisavljevic, V., Reljic, M., Cizek-Sajko, M., 2004.
 518 Developmental capacity of different morphological types of day
 519 5 human morulae and blastocysts. Reprod. Biomed. Online 8,
 520 687–694.
- Kresowik, J.D., Sparks, A.E., Van Voorhis, B.J., 2012. Clinical factors associated with live birth after single embryo transfer.
 Fertil. Steril. 98, 1152–1156.
- Land, J.A., Evers, J.L., 2003. Risks and complications in assisted reproduction techniques: Report of an ESHRE consensus meeting. Hum. Reprod. 18, 455–457.
- Martikainen, H., Tiitinen, A., Tomás, C., Tapanainen, J., Orava, M.,
 Tuomivaara, L., Vilska, S., Hydén-Granskog, C., Hovatta, O.the
 Finnish ET Study Group, 2001. One versus two embryo transfer
 after IVF and ICSI: a randomized study. Hum. Reprod. 16,
 1900–1903.
- McLernon, D.J., Harrild, K., Bergh, C., Davies, M.J., de Neubourg,
 D., Dumoulin, J.C., Gerris, J., Kremer, J.A., Martikainen, H.,
 Mol, B.W., Norman, R.J., Thurin-Kjellberg, A., Tiitinen, A., van
 Montfoort, A.P., van Peperstraten, A.M., Van Royen, E., Bhattacharya, S., 2010. Clinical effectiveness of elective single
 versus double embryo transfer: meta-analysis of individual
 patient data from randomised trials. BMJ 341, c6945.
- Pandian, Z., Bhattacharya, S., Ozturk, O., Serour, G., Templeton,
 A., 2009. Number of embryos for transfer following in-vitro
 fertilisation or intra-cytoplasmic sperm injection. Cochrane
 Database Syst. Rev., CD003416.
- Papanikolaou, E.G., Kolibianakis, E.M., Tournaye, H., Venetis, C.A.,
 Fatemi, H., Tarlatzis, B., Devroey, P., 2008. Live birth rates
 after transfer of equal number of blastocysts or cleavage-stage
 embryos in IVF. A systematic review and meta-analysis. Hum.
 Reprod. 23, 91–99.

- Papanikolaou, E.G., Camus, M., Kolibianakis, E.M., Van Landuyt, L., Van Steirteghem, A., Devroey, P., 2006. In vitro fertilization with single blastocyst-stage versus single cleavage-stage embryos. N. Engl. J. Med. 354, 1139–1146.
- Prados, F.J., Debrock, S., Lemmen, J.G., Agerholm, I., 2012. The cleavage stage embryo. Hum. Reprod. 27, i50–i71.
- Puissant, F., Van Rysselberge, M., Barlow, P., Deweze, J., Leroy, F., 1987. Embryo scoring as a prognostic tool in IVF treatment. Hum. Reprod. 2, 705–708.
- Racowsky, C., Combelles, C., Nurredin, A., Pan, Y., Finn, A., Miles, L., Gale, S., O'Leary, T., Jackson, K.V., 2003. Day 3 and Day 5 morphological predictors of embryo viability. Reprod. Biomed. 6, 323–331.
- Rehman, K.S., Bukulmez, O., Langley, M., Carr, B.R., Nackley, A.C., Doody, K.M., Doody, K.J., 2007. Late stages of embryo progression are a much better predictor of clinical pregnancy than early cleavage in intracytoplasmic sperm injection and in vitro fertilization cycles with blastocyst-stage transfer. Fertil. Steril. 87, 1041–1052.
- Richter, K.S., Harris, D.C., Daneshmand, S.T., Shapiro, B.S., 2001. Quantitative grading of a human blastocyst: optimal inner cell mass size and shape. Fertil. Steril. 76, 1157–1167.
- Shapiro, B.S., Daneshmand, S.T., Garner, F.C., Aguirre, M., Thomas, S., 2008. Large blastocyst diameter, early blastulation, and low preovulatory serum progesterone are dominant predictors of clinical pregnancy in fresh autologous cycles. Fertil. Steril. 90, 302–309.
- Terriou, P., Sapin, C., Giorgetti, C., Hans, E., Spach, J.L., Roulier, R., 2001. Embryo score is a better predictor of pregnancy than the number of transferred embryos or female age. Fertil. Steril. 75, 525–531.
- Thurin, A., Hardarson, T., Hausken, J., Jablonowska, B., Lundin, K., Pinborg, A., Bergh, C., 2005. Predictors of ongoing implantation in IVF in a good prognosis group of patients. Hum. Reprod. 20, 1876–1880.
- Thurin, A., Hausken, J., Hillensjö, T., Jablonowska, B., Pinborg, A.,
 Strandell, A., Bergh, C., 2004. Elective single embryo transfer versus double-embryo transfer in in vitro fertilization. N. Engl. J. Med. 351, 2392–2402.
- Van Royen, E., Mangelschots, K., De Neubourg, D., Valkenburg, M.,
 Van de Meerssche, M., Ryckaert, G., Eestermans, W., Gerris, J.,
 1999. Characterization of a top quality embryo, a step towards single-embryo transfer. Hum. Reprod. 14, 2345–2349.
- Wilson, M., Hartke, K., Kiehl, M., Rodgers, J., Brabec, C., Lyles, R., 2004. Transfer of blastocysts and morulae on day 5. Fertil. Steril. 82, 327–333.
- Yoon, H.J., Yoon, S.H., Son, W.Y., Im, K.S., Lim, J.H., 2001. High implantation and pregnancy rates with transfer of human hatching day 6 blastocysts. Fertil. Steril. 75, 832–833.
- Ziebe, S., Petersen, K., Lindenberg, S., Andersen, A.G., Gabrielsen,
 A., Nyboe Andersen, A., 1997. Embryo morphology or cleavage stage: how to select the best embryos for transfer after in-vitro fertilization. Hum. Reprod. 12, 1545–1549.
 600

Declaration: The authors report no financial or commercial 601 conflicts of interest. 602

Received 1 March 2013; refereed 5 July 2013; accepted 9 July 2013.